LOCATION OF FUNGI IN PUMPKIN SEED

Nasreen Sultana

Crop Diseases Research Institute, Pakistan Agricultural Research Council, Karachi University Campus, Karachi-75270, Pakistan

(Received 29 June 2001; accepted 11 July 2002)

Key words: Pumpkin seeds, Fungi, Location.

Pumpkin (Cucurbita pepo L.) is an important and widely cultivated cucurbitaceous vegetable crop in Pakistan. Over forty-five seed-borne fungi have been reported to be associated with pumpkin seeds in Pakistan (Yunis & Kauser, 1966; Sheikh 1990; Sultana et al 1992; Ahmed et al 1993). Macrophomina phaseolina, Botryodiplodia theobromae, Fusarium equiseti, F. semitectum, F. solani and F. oxysporum were found in high frequencies in cucurbits seed (Maholay

& Sohi 1982; Maholay 1988, 1989; Sheikh 1990; Sultana et al 1992). A knowledge of the exact location of seed-borne fungi in different seed components will be helpful in understanding their seed to plant transmission, effect in seed germination, disease cycle and control strategy. Studies were therefore carried out to locate the fungi on different components of pumpkin seed.

Six samples of pumpkin seeds were collected from market and farmer field in Pakistan, used to study the location of seedborne fungi. For determining the location of fungi in seeds, they were washed and soaked in distilled water for 2 hrs and then aseptically dissected to separate the different components viz. seed coat, tegmen, cotyledon and embryo. Dissected seed parts were surface disinfected with 0.5% NaOCl solution and placed on blotters in petri dishes (Du-Hyunglee *et al* 1984). The dishes were incubated at 24°C for 7 days and fungi, growing on different parts, were identified by using references of Raper & Fennel (1965),

Table 1

Percentage recovery of fungi from different seed parts of 6 seed samples of pumpkin

Name of fungi	Seed coat		Tegmen		Cotyledons		Embryo	
	SI	PI	SI	PI	SI	PI	SI	PI
Alternaria alternata	3	2.30±0.51				-	_	-
Aspergillus flavus	4	4.25±0.59	1	1.00±0.00	3	1.67±0.38	-	-
A. niger	:4	3,75±0.31	-		-	-		
A. terreus	2	3.50±1.06	1	1.00 ± 0.00		-	-	-
A. wentii	3	2.50±0.23	1	1.00±0.00	2	1.50 ± 0.35	-	-
Botryodiplodia theobromae	2	37.00±23.33	2	28.50±16.61	1	3.00 ± 0.00	1	1.00 ± 0.00
Chaetomium funicola	1	5.00±0.00	1	1.00±0.00			-	-
C. globossum	1	6.00±0.00	-		-		-	
C. olivaceum	1	4.00±0.00	1	1.00±0.00			-	
Cephaliophora irregularis	1	1.00±0.00	5.6	F.				
Curvularia clavata	1	1.00±0.00	1	1.00±0.00	+			
C. lunata	2	1.50±0.35	-	+:				
Drechslera halodes	4	2.67±0.14	2	1.50±0.35	1	2.00±0.00		
D. spicifera	3	2.00±0.33				-	-	-
Fusarium moniliforme	4	4.00±0.79	2	2.50±1.06	- 2	1.50±0.35	1	1.00 ± 0.00
F. oxysporum	5	14.60±2.02	- 5	7.00 ± 0.80	- 4	6.40±0.89	3	2.20±0.82
F. semitectum	4	2.75±0.37	2	1.00±0.00	2	2.00±0.00	-	-
F. solani	3	4.00±0.66	2	2.00±0.00	2	1.00 ± 0.00	1	1.00 ± 0.00
Macrophomina phaseolina	3	28.30±15.20	3	30.00±13.57	1	9:00±0.00	1	5.00±0.00
Myrothecium roridum	2	2.50±0.35		-			-	
M. verrucaria	1	6.00±0.00	7.00	-		-		
Penicillium purpurogenum	3	3.00±0.57	2	1.50±0.35	3	1.00±0.00	-	-
Stachybotrys atra	2	2.00±0.70	ii e	-		-	-	-
Sordaria tetraspora	1	3.00±0.00						
S. fumicola	1	4.00±0.00						
Ulocladium atrum	1	2.00±0.00		_	-		-	-

SI, Number of samples infected; PI, Percentage of infected seed ± standard error.

212 Short Communication

Booth (1971), Ellis (1971), Barnett & Hunter (1972), Nelson *et al* (1983).

The results indicated that most of the fungi were located on seed coat followed by tegmen (14), cotyledons (10) and embryo (5) with an average infection of 5.87, 5.71, 2.71 and 2.04% respectively. A total of 26 fungal species had been isolated Table 1. B. theobromae, F. moniliforme, F. oxysporum, F. solani and M. phaseolina were recovered from each component of the seed. The inoculum level decreased with the depth. Mathur et al (1975) and Sultana et al (1988) found that infection of F. moniliforme was more conveniently detected in the endosperm than seed coat and embryo whereas Bhutta et al (1996) and Dawer & Ghaffar (1998) found equal infection level of M. phaseolina in the pericarp, endosperm and embryo. In the present study B. theobromae and M. phaseolina were having high infection level in seed coat and tegmen while it was recovered from the embryo of only one seed samples.

Aspergillus flavus, A. wentii, Drechslera halodes, Fusarium semitectum and Penicillium purpurogenum were showing high level of infection on seed coat and cotyledons as compared to tegmens. High infection and deep penetration of the fungus in endosperm/cotyledons and embryo are considered as the major factors in germination failure and pre-emergence mortality (Raut 1983; Sultana et al 1994). Aspergillus terreus, Chaetomium fumicola, C. olivaceum and Curvularia clavata were having infection in seed coat and tegmen while rest of the fungi viz. Alternaria alternata, Aspergillus niger, Chaetomium globossum, Cephaliphora irregularis, Curvularia lunata, Drechslera spicifera, Myrothecium roridum, M. vernaria, Stachybotrys atra, Sordaria tetrapora, S. fumicola and Ulocladium atrum were found in seed coat only.

It is evident from these studies that the pathogenic fungi like *B. theobromae*, *F. moniliforme*, *F. oxysporum*, *F. solani* and *M. phaseolina* having both extra and intra embryonal infection affect seed quality immensely. Saprophytic as well as pathogenic fungi like *Aspergillus flavus*, *A. wentii*, *Drechslera halodes* and *Fusarium semitectum* occur not only in the seed coat but also in inner part of the seed of pumpkin, which impaired the seed quality.

References

- Ahmad I, Iftikhar S, Bhutta A R 1993 Seed-Borne Microorganism in Pakistan, Pakistan Agricultural Research Council, Islamabad. pp 32.
- Barnett H L, Hunter B B 1972 *Illustrated Genera of Imperfect Fungi*, Burgess Publishing Co. Minnesota. pp 241.

Bhutta A R, Bhatti M H R, Nizamani M S, Ahmad I 1996 Location and seed to plant transmission of pathogens in sunflower. *Pak J Phytopathol* 8(2) 172-176.

- Booth C 1971 *The Genus Fusarium*. Commonwealth Mycological Inst, Kew, Surrey, England. pp 237.
- Dawar S, Ghaffar A 1990 Location of Fungi in Sunflower Seed. *Pak J Bot* 23(2) 117-120.
- Du-Hyunglee, Mathur S B, Neergaard P 1984 Detection and location of seedborne inoculum of *Didymella bryomae* and its transmission in seedlings of cucumber and pumpkin. *Phytopath Z* **109** 301-308.
- Ellis M B 1971 Dematiaceous Hyphomycetes. CMI, Kew, Surrey, England, pp 608.
- Maholay M N, Sohi S H 1982 *Botryodiplodia* seed rot of bottlegourd and squash. *Indian J Mycol & Pl Pathol* 12 32-36.
- Maholay M N 1988 Seedborne diseases of cucurbits Musk-melon (*Cucumis melo* L.) Seed & Farms 14 11-12.
- Maholay M N 1989 Seedborne diseases of cucurbits. III Bottlegourd (*Lageneria siceraria* (Mol., Standl.). Seed & Farms 15 30-31.
- Mathur S K, Mathur S B, Neergaard P 1975 Detection of seedborne fungi in sorghum and location of *Fusarium* moniliforme in the seed. Seed Sci and Technol 3 683-690.
- Nelson P E, Toussoun T A, Marasas W F U 1983 Fusarium Species An Illustrated Manual for Identification. The Pennsylvania State University Press. pp 193.
- Raper K E, Fennel D I 1965 *The Genus Aspergillus*. Williams and Wilkins Co. Baltimore, pp 686.
- Raut JG 1983 Transmission of seedborne Macrophomina phaseolina in sunflower. Seed Sci & Technol 11 807-814.
- Sheikh AW 1990 Seedborne pathogen of vegetables crop grown in Pakistan. Summaries of Research Project 1967-1968. DGISP, Denmark.
- Sultana N, Khan SAJ, Khanzada AK 1988 Detection and location of seedborne fungi in sorghum seed. *Pak J Bot* **20** 239-244.
- Sultana N, Khanzada AK, Ghaffar A 1992 Seedborne fungi associated with pumpkin (*Cucurbita pepo L.*). Proceedings of National Symposium Status of Plant Pathology in Pakistan. Dept. Botany, University of Karachi, 1991. pp 117-123.
- Sultana N, Khanzada AK, Ghaffar A 1994 Location of Macrophomina phaseolina in seeds of pumpkin and development of charcoal rot disease. Pak J Bot 26(1) 177-180.
- Yunis M, Kausar AG 1966 Prevalence and control of fungi occurring on seeds of certain summer vegetables. *Pak J Agri Sci* 3 7-15.